

16S rDNA Typing of *Salmonella* Typhi Strains from Different Geographical Locations in Sumba Island East, Nusa Tenggara, Indonesia

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A total of thirteen isolates representative of *Salmonella* Typhi from different geographical locations in Sumba Island, East Nusa Tenggara, Indonesia were identified by 16S rDNA gene sequences. Bacterial DNA extraction was prepared by using a Purelink™ Genomic DNA kit. The bacterial DNA and control were amplified using the specific primers for *S. Typhi*. These 16S rDNA gene sequence data were aligned with the corresponding available *S. Typhi* sequence and the reference organisms from the family Enterobacteriaceae from NCBI database by using the CLUSTAL X software. Phylogenetic trees were generated using the PHYLIP software package and the matrix of nucleotide similarity and nucleotide difference were generated by using the PHYDIT software. The results from the 16S rDNA analysis showed that the degree of similarity within these strains ranged from 99.13-100%. The percentage of sequence similarity between *S. Typhi* strains was very high (>99 %). Molecular phylogenetic analysis showed that all of the isolates formed a new center of diversity with *S. Typhi* ATCC 19430^T as a reference strain. Based on these results, all of the tested strains belonged to species of *S. Typhi* suggested by their relatedness with the type strain of *S. Typhi* ATCC 19430^T.

Key words: 16S rDNA, phylogenetic analysis, *Salmonella* Typhi, typing

Sebanyak 13 isolat *Salmonella* Typhi yang mewakili berbagai wilayah geografis di pulau Sumba, Nusa Tenggara Timur, Indonesia diidentifikasi berdasarkan sekuen gen 16S rDNA. DNA bakteri diekstraksi sesuai dengan petunjuk dari Purelink™ Genomic DNA kit. DNA bakteri dan kontrol diamplifikasi menggunakan primer spesifik untuk *S. Typhi*. Urutan basa sekuen gen 16S rDNA dianalisis bersama dengan sekuen 16S rDNA *S. Typhi* dan anggota famili Enterobacteriaceae yang diperoleh dari database NCBI menggunakan program ClustalX. Pohon filogeni dikonstruksi dengan menggunakan program PHYLIP sedangkan matriks similaritas dan perbedaan nukleotida dianalisis dengan menggunakan program PHYDIT. Hasil analisis sekuen 16S rDNA menunjukkan bahwa kemiripan di antara isolat *S. Typhi* berkisar antara 99,13-100%. Persentase kemiripan sekuen di antara isolat *S. Typhi* sangat tinggi (>99%). Analisis filogeni menunjukkan bahwa semua isolat membentuk pusat keanekaragaman baru dengan isolat standard *S. Typhi* ATCC 19430^T. Berdasarkan hasil tersebut, dapat disimpulkan bahwa semua isolat yang diuji tergolong dalam spesies *S. Typhi* yang ditunjukkan dari kedekatan hubungan kekerabatan dengan isolat standard *S. Typhi* ATCC 19430^T.

Kata kunci: 16S rDNA, analisis filogenetik, *Salmonella* Typhi, typing

Molecular techniques have become increasingly popular and potentially useful tool for the classification and identification of bacterial strains in most bacterial genera. One of which is 16S rDNA gene sequence analysis. The 16S rDNA gene is highly conserved within a species and among species of the same genus, and therefore it can be used as a reference for the speciation of bacteria (Woo *et al.* 2000). This method plays an important role in the identification of bacterial pathogens, which is useful not only for diagnosis, but also for phylogenetic classification. Most *Salmonella* strains are identified serologically as having O (somatic) and H (flagellar) antigens. There are, however, often serological cross-reaction between strains that phylogenetic classification is important to distinguish them from one another (Woo *et al.* 2001).

Currently there are three known species in the *Salmonella* genus, *Salmonella enterica*, *Salmonella bongori*, and *Salmonella subterranea* (Shelobolina *et al.* 2004). *Salmonella enterica* comprises of six subspecies. They are *S. enterica* subsp. *enterica* (subsp. I), *S. enterica* subsp. *salamae* (subsp. II), *S. enterica* subsp. *arizonae* (IIIa subsp.), *S. enterica* subsp. *diarizonae* (subsp. IIIa), *S. enterica* subsp. *houtenae* (subsp. IV), and *S. enterica* subsp. *indica* (subsp. VI). *Salmonella bongori* was originally classified as *S. enterica* subspecies V. The majority of *S. enterica* subsp. *enterica* (subsp. I) cause most infections in humans and warm-blooded animals (Truper 2005; Tajbakhsh *et al.* 2011).

Salmonella Typhi, the agent of typhoid fever, is included in subspecies I: *S. enterica* subsp. *enterica* serotype Typhi (*S. enterica* serovar Typhi; *Salmonella*

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Typhi). Typhoid fever is most prevalent in tropical areas, including Indonesia (Moehario 2009). In Sumba Island, East Nusa Tenggara especially in Southwest Sumba District over 197 cases or 725 infection/100.000 inhabitants were recorded in the database of the Karitas Hospital in 2006 (Amarantini *et al.* 2009). This number was higher than the average infection cases in rural areas in Indonesia, and it was nearly the same as the average cases in urban areas (810/100.000) according to World Health Organization (WHO) in 2003. These numbers suggested that the area was a good niche for fast growing microorganisms. Analysis of various *S. Typhi* strains using numerical systematic method showed that there were a big diversities in the use of the carbon sources by the *S. Typhi* isolates in these region (Amarantini *et al.* 2009). In addition to obtaining accurate data and strong discriminative ability to distinguish the strains, this study was aimed to identify and unravel the diversities of *S. Typhi* isolates from typhoid fever patients using molecular phylogenetic approach based on 16S rDNA gene sequences.

MATERIALS AND METHODS

Bacterial Strains. Thirteen isolates used in this research were isolated from the blood cultures of typhoid patients in Sumba Island. They came from different geographical regions in Karitas Hospital in Weetabula, a private clinic in Elopada Subdistrict in Southwest Sumba District, and Lendemoripa Hospital in Waikabubak in West Sumba District. Specimen collection methods were described in the journal article published previously (Amarantini *et al.* 2009). These isolates were identified using microbiological standard methods. All cultures were screened in Chromocult Coliform Agar (CCA). Typical colonies appear white and transparent due to the lack of β -galactosidase and β -glucuronidase enzymes. These colonies were confirmed using Triple Sugar Iron Agar (TSIA) (WHO 2003). All cultures were grown at 30 °C for 24 h on Brain Heart Infusion (BHI) agar before used.

Extraction of Bacterial DNA, PCR Amplification, and DNA Sequencing. Bacterial DNA was extracted in accordance with the protocol's instructions using a Purelink™ Genomic DNA Mini Kit (Invitrogen K1820-00). The bacterial DNA and control were amplified with 25 pmol primers (1stBase Custom Oligos FBCO) and PCR SuperMix (illustra™ PuRe *Taq*™ Ready-To-Go™ PCR beads. PCR amplification for the 16S rDNA sequences showed bands of 428 bp, 484 bp, and 483 bp. These fragments were amplified using

primer R1 FR, R3 FR, and R5 FR respectively. The primers used to amplify these fragments are shown in Table 1 (Massi *et al.* 2005). The PCR mixtures were amplified for 40 cycles at 94 °C for 1 minute, 55 °C for 1 min, and 72 °C for 2 min, with a final extension at 72 °C for 10 min in automated Applied Biosystems GeneAmp PCR System 2400. An aliquot of 5 μ L of each amplified product was electrophoresed in 3.0% (w/v) agarose gel, with a 100 bp DNA Ladder (Gene Ruler™).

The PCR product was gel purified with a QIAquick PCR purification kit (QIAGEN, Hilden, Germany). The purified PCR product was sequenced with ABI Prism 3100-Avant Genetic Analyzer according to the manufacturer's instructions (Applied Biosystems, USA) using the same primers as used in PCR.

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Analysis and Alignment of 16S rDNA Nucleotide Sequences. The 16S rDNA nucleotide sequences were analyzed, edited and assembled with Finch TV 1.4.0 and DNA Baser sequence analysis software. Complete assembled sequences were aligned with the corresponding *S. Typhi* sequences retrieved from the NCBI database with CUSTAL X software (Thompson *et al.* 1997).

Construction of Phylogenetic Tree. Based on 16S rDNA nucleotide sequences, a phylogenetic tree was constructed with PHYLIP software package (Felsenstein 1993) with neighbor-joining algorithm (Saito and Nei 1987). The evolutionary distance matrix of the neighbor-joining method was generated according to the description from Jukes and Cantor (1969). The phylogenetic distances were obtained by adding only the values of the horizontal components. Eventually, the matrix of the nucleotide similarity and difference was generated with PHYDIT software (Chun 1999).

RESULTS

Thirteen isolates used in this research came from different locations; seven isolates from East Wewewa, three isolates from Kodi, two isolates from North Wewewa, and one isolate from Waikabubak. These isolates were selected to represent their geographical origins. They're mapped according to the infected patient's place of residence based on global positioning system (Fig 1).

Phylogenetic analysis results of 13 *S. Typhi* isolates based on 16S rDNA gene sequences are shown in Fig 2. The 16s rDNA gene sequences of these isolates were initially compared to those of the *Enterobacteriaceae* family reported in the Gene Bank with the accession

Omba Rade), HB 10 (isolated from Weedindi), HB11 (isolated from Durru Lodo), HB12 (isolated from Kongge), and HB13 (isolated from Elopada).

The second clade consisted of five strains, which were HB02 (isolated from Kampung Sawah Waikabubak),

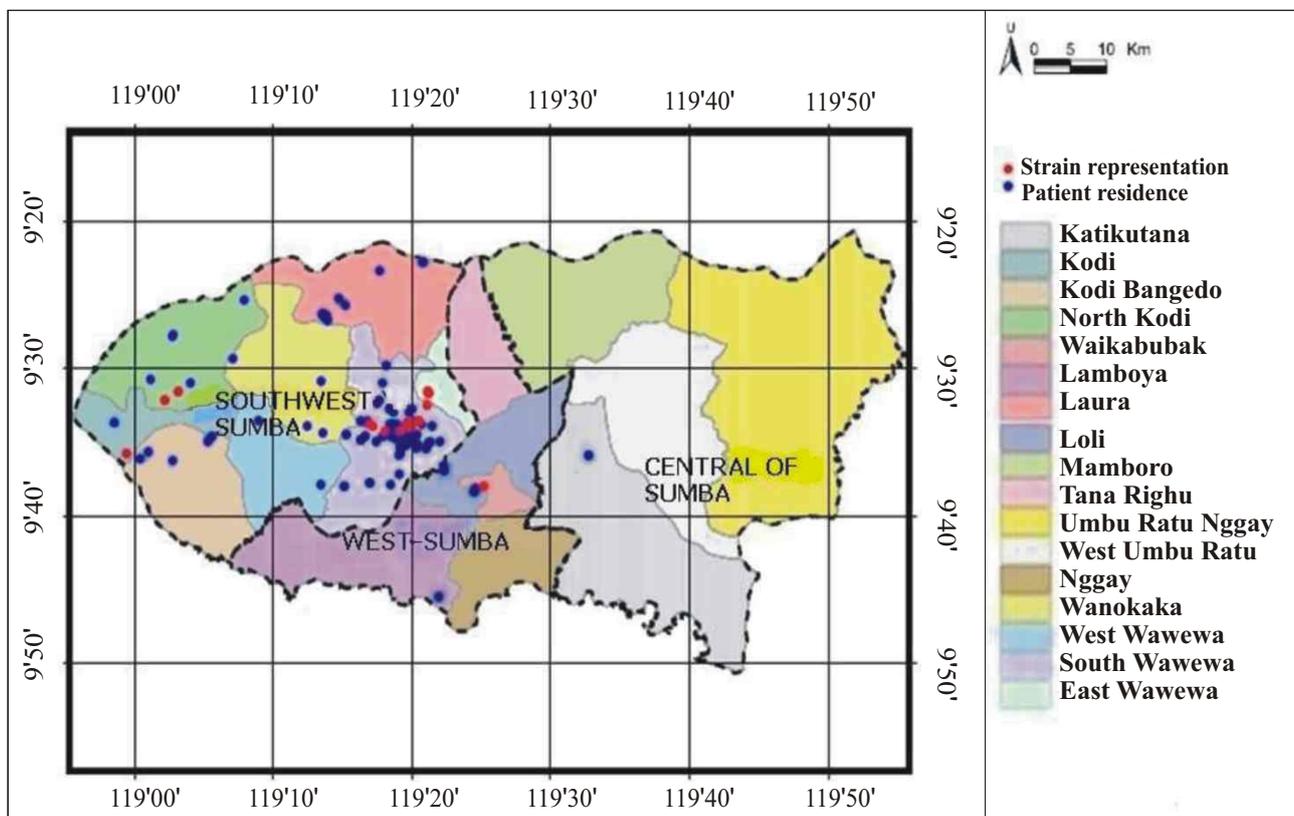


Fig 1 Distribution of typhoid fever patients and 13 strains representation from different geograophic location in Sumba Island, East Nusa Tenggara.

number Z47544 (*S. Typhi* ATCC 19430^T), AF029227 (*S. bongori* BR 1859), X80724.1 (*Escherichia coli* ATCC 25922), M59291 (*Citrobacter freundii* ATCC29935), M59160 (*Serratia marcescens*), M59149 (*Erwinia carotovora* ATCC 15713), X75275 (*Yersinia ruckeri* ATCC 29473), X75279 (*Yersinia intermedia* ER-3854), M59155 (*Hafnia alvei* ATCC 13337), X82248 (*Photobacter luminescens* DSM 3368), X82251 (*Xenorhabdus nematophilus* DSM 3370), X07652 (*Proteus vulgaris* IFAM 1731), and M59159 (*Plesiomonas shigelloides*)

The phylogenetic analysis showed 13 representative strains of *S. Typhi* originated from different locations in Sumba Island, East Nusa Tenggara, which were then divided into three clades. The first clade consisted of seven strains. One isolate (HB01) originated from Wailabubu, North Kodi Subdistrict and six isolates were obtained from East Wewewa Subdistrict, which were HB05 (isolated from Wanowitu), HB09 (isolated from

HB07 (isolated from Weerambo, East Wewewa), HB06 (isolated from Ombawawi North Wewewa), HB03 (isolated from Watubero Wailabubur North Kodi), and HB04 (isolated from Pakamutu, Bondokodi Kodi).

The third clade consisted of two strains. They were HB08 (isolated from Palekki/Mahaloko in North Wewewa Subdistrict) and *S. Typhi* ATCC 19430^T, which was the reference strain.

The 16S rDNA nucleotide similarity values (%) and the number of nucleotide differences among 13 *S. Typhi* isolates from infected patients in different geographical locations in Sumba Island and the reference strain *S. Typhi* ATCC 19430^T are shown in Table 1 and 2. The sequences of the 13 strains and *S. Typhi* ATCC 19430^T showed >99% similarity. It was also evident that strain HB10 was identical with the strain HB09, and the strain HB11 was identical with the strains HB09 and HB10 (Table 2). The two other strains (HB03 and HB04) were also identicals (Table 3).

Table 1 The primers used for PCR amplification of 16S rDNA gene sequence of *Salmonella* Typhi

Primer	Sequence	Nucleotide position
R1F	5' AGTTTGATCCTGGCTCAG 3'*	3-20 (AC: Z47544)
R1R	5' AGTACTTTACAACCCGAAGG 3'*	411-430 (AC: Z47544)
R3F	5' AAGTACTTTCAGCGGGGA 3'*	424-441 (AC: Z47544)
R3R	5' TTGAGTTTTAACCTTGCGG 3'*	898-916 (AC: Z47544)
R5F	5' AACTCAAATGAATTGACGG 3'*	901-919 (AC: Z47544)
R5R	5' AGGCCCGGGAACGTATTCAC 3'*	1364-1383 (AC: Z47544)

AC: GenBank accession no. *Reference: Massi *et al.* 2005

Table 2 16S rDNA similarity values (%) and the number of nucleotide differences between seven strains and the reference strains of *S. Typhi* ATCC 19430^T within the first clade

	HB09	HB10	HB11	HB12	HB01	HB13	HB05	<i>S. Typhi</i> ATCC 19430 ^T	<i>S. bongori</i> BR1859
HB09	---	0/1383	0/1381	7/1382	4/1383	1/1382	1/1381	5/1381	33/1377
HB10	100.00	---	0/1381	7/1381	3/1382	1/1382	1/1381	5/1381	33/1377
HB11	100.00	100.00	---	7/1381	3/1381	1/1381	1/1381	5/1381	33/1377
HB12	99.49	99.49	99.49	---	11/1382	8/1381	8/1381	12/1381	40/1377
HB01	99.71	99.78	99.78	99.20	---	4/1382	4/1381	8/1381	36/1377
HB13	99.93	99.93	99.93	99.42	99.71		2/1381	6/1381	34/1377
HB05	99.93	99.93	99.93	99.42	99.71	99.86	---	4/1381	32/1377
<i>S. Typhi</i> ATCC 19430 ^T	99.64	99.64	99.64	99.13	99.42	99.57	99.71	---	36/1497
<i>S. bongori</i> BR1859	97.60	97.60	97.60	97.10	97.39	97.53	97.68	97.60	---

Table 3 16S rRNA similarity values (%) and the number of nucleotide differences between six strains and the reference strains of *S. Typhi* ATCC 19430^T within the two and the third clades

	HB03	HB04	HB06	HB02	HB07	HB08	<i>S. Typhi</i> ATCC 19430 ^T	<i>S. bongori</i> BR1859
HB03	---	0/1384	5/1381	5/1381	6/1381	4/1381	7/1381	35/1377
HB04	100.00	---	5/1381	5/1381	6/1381	4/1381	7/1381	35/1377
HB06	99.64	99.64	---	4/1381	5/1381	3/1381	6/1381	34/1377
HB02	99.64	99.64	99.71	---	1/1382	3/1381	6/1381	34/1377
HB07	99.57	99.57	99.64	99.93	---	4/1381	7/1381	35/1377
HB08	99.71	99.71	99.78	99.78	99.71	---	3/1381	33/1377
<i>S. Typhi</i> ATCC 19430 ^T	99.49	99.49	99.57	99.57	99.49	99.78	---	36/1497
<i>S. bongori</i> BR1859	97.46	97.46	97.53	97.53	97.46	97.60	97.60	---

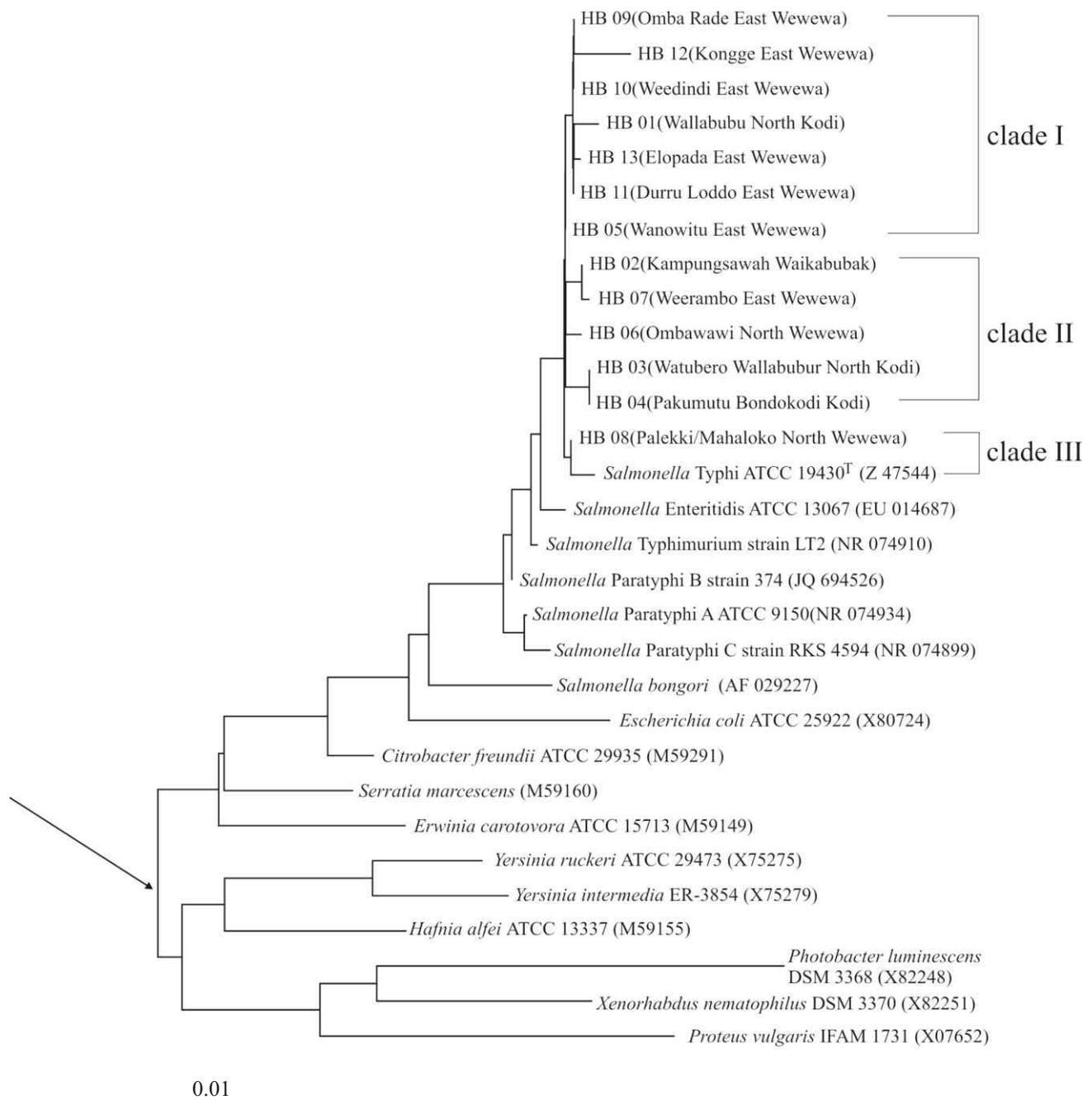


Fig 2 Neighbour-joining phylogeny tree constructed on the basis of 16S rDNA gene sequences showing relationship amongst the thirteen representatives of *S. Typhi* isolates from the different locations in Sumba Island, East Nusa Tenggara. The arrow indicates estimated root position of the tree as determined using *Plesiomonas shigelloides* (M59159) as an outgroup. Bar, 1 substitution per 100 nucleotides.

DISCUSSION

Phylogenetic analysis based on comparison of 16S rDNA nucleotide sequences of the 13 strains with the corresponding nucleotide sequence of available representatives of the genus *Salmonella* clearly showed that all of the tested strains form a new center of diversity with *S. Typhi* ATCC 19430^T (Fig 2). This result directly proved that all of the isolates belong to *S. Typhi* species because of their relationships with the

reference strain *S. Typhi* ATCC 19430^T. It was also shown that strains within the species have diverse 16S rDNA gene sequences. In fact, all of the strains fall into three clades. These clades demonstrated that genetic diversity of the tested strains could be unraveled using phylogenetic tree based on 16S rDNA sequences.

It is shown in phylogenetic tree that each clade is made up of strains from different geographical areas in the Sumba Island especially in Southwest Sumba District. The six tested strains within the first clade

were derived from East Wewewa Subdistrict, whereas the other isolates were derived from North Kodi Subdistrict. The second clade consisted of two strains originally from Kodi, one strain derived from Waikabubak, and the two strains derived from East Wewewa and North Wewewa. Based on the coordinate position of the infected patient's residences (Fig 1), we learned how the typhoid fever spreads among the inhabitants. Most of the patients lived in East Wewewa Subdistrict. It appears that the strains were distributed in finger pattern to nearby subdistrict such as North Wewewa, South Wewewa, West Wewewa, and then goes further out to the Kodi Bangedo, Kodi, and North Kodi. Therefore, these results indirectly showed that there were inter geographical distribution of the strains due to migration of people in this area.

Identification to the species level requires that the tested strains 16S rDNA sequence has 99% similarity with the sequence of the reference strain in GenBank (Drancourt *et al.* 2000). It is shown in this study that the 16S rDNA sequence of these 13 isolates had >99% similarity with the sequence of the closest strain in GenBank. Thus these isolates were identified as *S. Typhi*.

In terms of nucleotide similarity and nucleotide differences (Table 1), it was shown that the strain HB10 was identical with HB09, and the strain HB11 was identical with HB09 and HB10. However, this result did not entirely coherent with its phylogenetic relatedness based on phylogenetic tree. It is apparent in Fig 2 that the closest strain was found to be the strain HB12. Two tested strains, HB03, and HB 04 (Table 2), were fully congruent in their similarity values, nucleotide differences and phylogenetic analysis. It is especially interesting that the strain HB 08 from Palekki/Mahaloko North Wewewa was found to be closely related to *S. Typhi* ATCC 19430^T.

Based on analysis of housekeeping and rRNA genes, it has been understood that the genus *Salmonella* contains two lineages that had diverged considerably from each other during evolution. These lineages represent two distinct species, *S. enterica* and *S. bongori* (Baumler *et al.* 1998). Christensen *et al.* (1998) also demonstrated that the analysis of 16S rDNA sequences separated *S. bongori* from *S. enterica*, and these two species from the complex of *E. coli* and *Shigella* species. This corresponds to the topology of this phylogeny tree (Fig 2).

In summary, all the isolates were identified as *S. Typhi* species because of their association with the type strain of *S. Typhi* ATCC 19430^T. The fact that the 13 isolates belonging to *S. Typhi* species formed a new center of diversity within the 16S rDNA gene tree

indicated that these strains are indigenous from the Southwest Sumba District.

ACKNOWLEDGMENTS

This research was supported by The Directorate General of Higher Education, Department of National Education (Hibah Bersaing) 2012, contract no: 560.7/K5/KL/2012 date 10-02-2012. A special gratitude is given to Karitas Hospital in Weetabula in Southwest Sumba District and Lende Moripa Hospital in Waikabubak in West Sumba District East Nusa Tenggara for their assistance in collecting the samples. I should also thank Sr. Sili Bouka ADM-the Director of Karitas Hospital, dr. Loeta Lapoe Moekoe-the Director of Lende Moripa Hospital and all doctors of Karitas Hospital for their assistance during the research.

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