

# The Growth of Leptolyngbya HS-16 and HS-36 on 35 °C at Different Acidity

## NURUL RAKHMAYANTI, NINING BETAWATI PRIHANTINI\*

Department of Biology, Faculty of Mathematics and Natural Sciences, Universitas Indonesia, Kampus UI Depok 16424, Indonesia

Cyanobacteria are prokaryotic organisms belonging to the kingdom Eubacteria. Cyanobacteria can be found in hot spring. *Leptolyngbya* is one genus of cyanobacteria that can be found in hot spring. The observation of *Leptolyngbya* growth on temperature of 35 °C with initial pH variation had been done. The study was experimental trial. The study aimed to determine the best initial growth pH for *Leptolyngbya* HS (Hot Spring)-16 and HS-36. *Leptolyngbya* HS-16 was isolated from Pancar Mountain hot spring, while *Leptolyngbya* HS-36 was isolated from Maribaya hot spring. The acidity (pH) of Pancar Mountain and Maribaya hot spring was 7. Each strain was grown in Blue Green medium number 11 with variation of initial pH (6, 7, 8 and 9) and incubated at 35 °C. Parameters was wet biomass weight of *Leptolyngbya* in each strain. The results of 15 days observation showed that the best initial pH for growing *Leptolyngbya* HS-16 is 7, while *Leptolyngbya* HS-36 is 9. From this study it could be seen that *Leptolyngbya* HS-16 and HS-36 could be cultured with alkaline condition.

Key words: hot spring, Leptolyngbya, pH

Cyanobacteria merupakan organisme prokariotik yang berasal dari kingdom *Eubacteria*. Cyanobacteria dapat ditemukan pada sumber air panas. Salah satu genus dari cyanobacteria yang ditemukan dalam sumber air panas, yaitu *Leptolyngbya*. Pengamatan pertumbuhan *Leptolyngbya* pada suhu 35 °C dengan variasi pH awal telah dilakukan. Penelitian ini merupakan penelitian eksperimental. Penelitian ini bertujuan untuk mengetahui pH pertumbuhan awal terbaik untuk *Leptolyngbya* HS-16 dan HS-36. *Leptolyngbya* HS-16 diisolasi dari sumber air panas di gunung Pancar, sedangkan *Leptolyngbya* HS-36 diisolasi dari sumber air panas di Maribaya. Derajat keasaman (pH) air dari sumber air panas di gunung Pancar dan Maribaya adalah 7. Masing-masing strain ditumbuhkan pada medium Blue Green nomor 11 dengan variasi pH awal (6, 7, 8 dan 9) dan diinkubasi pada suhu 35 °C. Parameter yang diteliti adalah berat basah biomassa *Leptolyngbya* pada masing-masing strain. Pengamatan dilakukan selama 15 hari dengan 11 sampling. Hasil pengamatan 15 hari menunjukkan bahwa pH awal terbaik untuk pertumbuhan *Leptolyngbya* HS-16 adalah 7, sedangkan *Leptolyngbya* HS-36 adalah 9. Berdasarkan penelitian dapat diketahui bahwa *Leptolyngbya* HS-16 dan HS-36 dapat dibiakkan dengan kondisi alkalin.

Kata kunci: Leptolyngbya, pH, sumber air panas

Cyanobacteria are prokaryotic organisms belonging to the kingdom Eubacteria (Van den hoek 2002). These organisms has photosynthetic apparatus that plays role in producing energy (Scholnick *et al.* 2006). Cyanobacteria has photosynthetic pigment named phycobilin. These organisms stores glycogen as its food storage (Markou *et al.* 2014).

The growth phases of cyanobacteria are same with other microorganisms. The growth curve of population in microorganisms known as exponential growth. Exponential growth in microorganisms consist of lag phase, exponential phase, stationary phase and death phase. Lag phase occur when microorganisms has been innocculated into a new medium and adaptation with new environmental with different condition of their habitat. Exponential phase happened when microorganism could adapted with their new

environmental condition and their population increased twice of their first population. The population of microorganisms increased because they could used source of their new environmental. Stationary phase occurred after exponential phase. Stasioner phase is the phase that microorganisms could not do doubling cell. This phase occurred because the nutrients present in the growth medium are insufficient for doubling cell and accumulation of the metabolic waste of the microorganisms. Death phase will occured after stationer phase (Madigan *et al.* 2015).

Cyanobacteria can be found in soil, rocks and waters (Bold *et al.* 1978). Another source where cyanobacteria can be found is hot spring. Maribaya hot spring and Pancar mountain hot spring are the example of source whereas cyanobacteria can be found. Maribaya hot spring has pH 6 to 7, while Pancar Mountain hot spring has pH 7. *Leptolyngbya* is one genus of cyanobacteria that can be found in Maribaya and Pancar Mountain hot spring (Prihantini 2015).

<sup>\*</sup>Corresponding author: Phone: +62-81297776638; Fax: +62-21-7270012; Email: nining@ui.ac.id

70 RAKHMAYANTI ET AL. Microbiol Indones

Leptolyngbya have filamentous as the form of the colony. Characteristic of these organisms have a thin filament with 0.5 to 3.5 μm wide with simple trichome, and some species have sheath (Komarek 2007). Leptolyngbya can be found in environmental condition with pH 7 to 8.5 (Olsson-francis et al. 2012)

Leptolyngbya have many benefit for our life. It could be seen Leptolyngbya could produce lipid as biofuel feedstock, produce secondary metabolites as antibiotic, and could react as bioremediator in dairy waste (Abazari et al 2012; Beetul, 2014; Khemka et al 2015). In order to beneficial of Leptolyngbya, it is important to cultivated *Leptolyngbya*. The acidity (pH) in the environment can affect the growth rate of cyanobacteria especially Leptolyngbya. Cyanobacteria can also live in environmental conditions with a wide range of pH, but some species are sensitive to acidic conditions (Gerloff-Elias et al, 2005). Cyanobacteria could barely find in freshwater with range of pH 4 to 5 (Bold et al. 1978). Variation of initial pH in growth medium will affect the growth of cyanobacteria. The Leptolyngbya HS-16 and HS-36's best initial growth pH have not known yet. The aim of this study is to determine the best initial growth pH for Leptolyngbya HS-16 and HS-36 in Blue Green number 11 medium (BG-11).

#### **MATERIALS AND METHODS**

Microorganisms and Growth Medium. The microorganisms used in this study were cyanobacteria genus *Leptolyngbya* strain HS-16 and HS-36. *Leptolyngbya* HS-16 was isolated from Pancar Mountain hot spring, while *Leptolyngbya* HS-36 was isolated from Maribaya hot spring. Those strains were grown in Blue Green number 11 medium/ BG-11 (NIES 2007) with variations of pH value 6, 7, 8 and 9. The BG-11 medium were made as reported by Prihantini (2015).

Cyanobacteria Cultivation in BG-11 Medium. The first step of cyanobacteria cultivation was inoculated 30 mg biomass of each strain into 100 mL growth medium in 250 mL Erlenmeyer flask. Before inoculation of cyanobacteria into medium, the medium had been adjusted the variation of pH value into 6, 7, 8 and 9. The treatment of variation pH value was repeated twice in each strain. Those strain were incubated at temperature 35 °C.

Measurement the weight of wet biomass Leptolyngbya HS-16 and HS-36. Measurement of *Leptolyngbya* HS-16 and HS-36 biomass were done in

15 days with 11 times of sampling. Sixteen of sterile eppendorf tube 2 mL was measured at analytical measurement tool. Biomass of those strain were taken aseptically with sterile micropipet amount of 2 mL. Eppendorf tube with biomass of those strain inside were centrifuged with Biofuge Primo R machine in room temperature for 10 minute (in 10.000 rpm). The supernatant of those strain were taken out and wet biomass weight were measured with analytical measurement tool. The growth curves were made by comparrasion between wet biomass weight as the ordinate axis Y with observed time as absisca X. The growth curves were made by Microsoft Excel.

#### RESULTS

The study of growth *Leptolyngbya* HS-16 and HS-36 had been done. It took 15 days with 11 times of sampling. The result produced growth curve of *Leptolyngbya* HS-16 and HS-36. The growth curve showed the growth of both strains in the adaptation stage to the pH condition of the medium. It could be seen at the growth curve of *Leptolyngbya* HS-16 and HS-36, which each strain produce the growth curve unstable. The age of inoculum that used in this study were 5 months old.

Macroscopic observation of *Leptolyngbya* were observed. Color appereance of *Leptolyngbya* based on Faber Castle standard color. Color appereance of *Leptolyngbya* HS-16 was emerald green, while *Leptolyngbya* HS-36 was brown ochre at day 0 (t0). At day-15 (t15), the color appereance of *Leptolyngbya* HS-16 in pH 9 was changed from emerald green into apple green, while *Leptolyngbya* HS-36 in pH 6, 7 & 8 were changed from brown ochre into apple green.

The average of wet weight biomass of *Leptolyngbya* HS-16 and HS-36 are shown on Table 1. The growth curve of *Leptolyngbya* HS-16 shown on Figure 3, while the growth curve of *Leptolyngbya* HS-36 shown on Figure 4. The growth curve of *Leptolyngbya* HS-16 and HS-36 were made based on their wet weight of biomass.

After 15th day of incubation, both *Leptolyngbya* were able to grow on initial pH 6 medium, with wet weight 0.0295 g L<sup>-1</sup> for HS-16 and 0.02905 g L<sup>-1</sup> for HS-36. The growth curve of those strain in initial pH 6 were slightly rise at 2<sup>th</sup> until 10<sup>th</sup> day, and drasticaly increased at 13<sup>th</sup> until 14<sup>th</sup> day, but decreased at 15<sup>th</sup> day. On the other medium with initial pH 7, *Leptolyngbya* HS-16 were able to grow and produced maximum amount of wet weight than other initial pH, but

Volume 12, 2018 Microbiol Indones 71

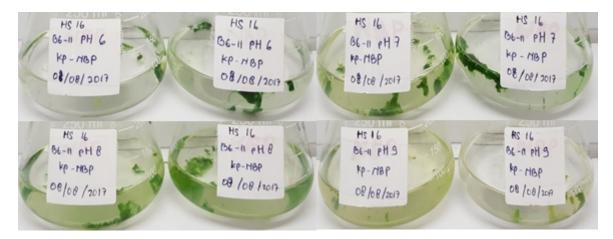


Fig 1 The color appereance of *Leptolyngbya* HS-16 at day-15 (t15).

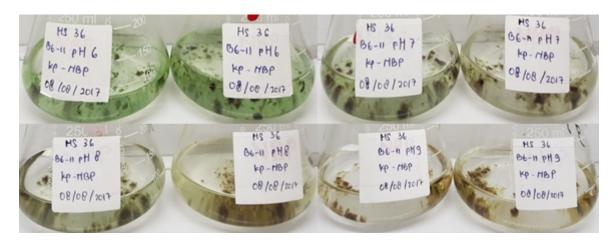


Fig 2 The color appereance of *Leptolyngbya* HS-36 at day-15 (t15).

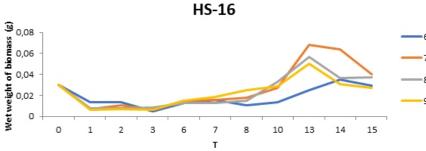


Fig 3 The growth curve of *Leptolyngbya* HS-16.

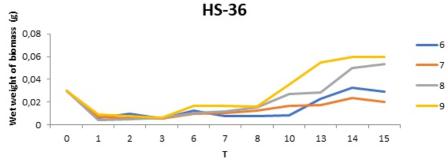


Fig 4 The growth curve of *Leptolyngbya* HS-36.

72 RAKHMAYANTI ET AL. Microbiol Indones

	Wet weight of biomass (♣ <sup>-1</sup> )							
T	HS-16				HS-36			
	6	7	8	9	6	7	8	9
0	0.03	0.03	0.03	0.03	0.03	0.03	0.03	0.03
1	0.01375	0.00685	0.00755	0.00645	0.00615	0.0069	0.0046	0.00925
2	0.01345	0.01075	0.00755	0.00705	0.0101	0.00545	0.0052	0.00745
3	0.00445	0.00655	0.00835	0.0061	0.00555	0.0058	0.0064	0.00645
6	0.01305	0.0151	0.01245	0.01515	0.0129	0.01	0.01075	0.0166
7	0.01535	0.01555	0.0126	0.0188	0.0077	0.0108	0.01205	0.01705
8	0.0107	0.0179	0.01465	0.02485	0.00755	0.0127	0.01525	0.0163
10	0.0132	0.02695	0.033	0.0285	0.0086	0.01675	0.02685	0.03555
13	0.02525	0.0681	0.05635	0.05005	0.02285	0.01765	0.0284	0.0548
14	0.0348	0.06415	0.0362	0.031	0.0325	0.02345	0.04965	0.0596
15	0.0295	0.0404	0.03725	0.02735	0.02905	0.01995	0.05345	0.05995

Table 1 The average of wet weight *Leptolyngbya* HS-16 and HS-36 (g L<sup>-1</sup>)

Leptolyngbya HS-36 produced minimum amount of wet weight than other initial pH. The wet weight of Leptolyngbya HS-36 was 0.0404 g L<sup>-1</sup> and Leptolyngbya HS-36 was 0.01995 g L<sup>-1</sup>. The growth curve of Leptolyngbya HS-16 in initial pH 7 was decreased at 2th until 4th day then increased at 3th until 13<sup>th</sup> day and decreased at 14<sup>th</sup> until 15<sup>th</sup> day, while Leptolyngbya HS-36 was increased constantly at 1th until 14<sup>th</sup> day and decreased at 15<sup>th</sup> day. Wet weight of Leptolyngbya HS-16 and HS-36 in growth medium with initial pH 8 were  $0.03725 \text{ g L}^{-1}$  and  $0.05345 \text{ g L}^{-1}$ . Both of those strain well adapted in medium with initial growth pH 8, especially Leptolyngbya HS-36. Leptolyngbya HS-16 was increased at 1th until 13th day then slightly decreased at 14th until 15th day, while Leptolyngbya HS-36 was increased at 1th until 15th day. Wet weight of Leptolyngbya HS-16 and HS-36 in growth medium with initial pH 9 were 0,02735 g L<sup>-1</sup> and 0,05995 g L<sup>-1</sup>. Leptolyngbya HS-36 produced maximum amount of wet weight than other initial pH. Leptolyngbya HS-16 was decreased at 1<sup>th</sup> until 3<sup>th</sup> day then increased at 6<sup>th</sup> until 13<sup>th</sup> day and decreased at 14<sup>th</sup> until 15th day, while Leptolyngbya HS-36 was decreased at 1<sup>th</sup> until 3<sup>th</sup> day then increased at 6<sup>th</sup> day, but slightly decreased again at 7th until 8th day and increased again until 15<sup>th</sup> day.

## **DISCUSSION**

Based on the result of this experiment, the color of *Leptolyngbya* HS-16 changed on growth medium with

initial pH 9, and *Leptolyngbya* HS-36 changed on growth medium with initial pH 6, 7, and 8. It hapened because of their physiology adaptation mechanism in new environmental. The physiological adaptation caused the alteration phycobilin content (Muster *et al.* 1983). As we can see on the growth curve, both strains still in lag phase on growth medium with initial pH 6, 7, 8, and 9. It proven that the curve still in unstable stage. Lag phase sometimes could be the longest phase for some microorganisms because in this phase, microorganisms must adapt with new environmental conditions like new source of nutrient, pH and temperature (Hogg 2005).

Both of these strains were able to grow in initial growth pH 6 medium, but not as good as in alkaline condition. Leptolyngbya were often be found in neutral to alkaline condition (Madigan et al. 2015). Leptolyngbya HS-16 had the most average in growth medium with pH 7 than other pH. It could be Leptolyngbya HS-16 was already adapted in growth medium with pH 7, because Leptolyngbya HS-16 was isolated from Pancar mountain, which has pH 7 (Prihantini 2015). Leptolyngbya HS-16 was well adapted in medium growth with pH 7, which coud be grouped into neutrophile organisms (Madigan et al. 2015). Leptolyngbya HS-36 were well adapted in growth medium with initial pH 8 and 9, especially Leptolyngbya HS-36 had the most average in growth medium with initial pH 9. It happened because Leptolyngbya were often found in environmental with pH 8 (Olsson-francis et al. 2012). It could be also that

Volume 12, 2018 Microbiol Indones 73

Leptolyngbya was alkalophile, which microorganisms that could live with pH 8 to 10 (Madigan et al. 2015). Those strains were decreased at 15<sup>th</sup> day of observation. It could be happened because the nutrients in the growth medium had been reduced. The nutrients in the growth medium used those strain for metabolisms to their growth (Madigan et al. 2015). Based on the observation of the 15<sup>th</sup> day and the discuccion that had been done, the best growth of Leptolyngbya HS-16 was on growth medium with initial pH 7, while the best growth of Leptolyngbya HS-36 was on growth medium with initial pH 9.

## **ACKNOWLEDGMENT**

This work was fully funded by Hibah Publikasi Internasional Terindeks untuk Tugas Akhir Mahasiswa (PITTA) 2017 to Nining Betawati Prihantini, grant no. 667/UN2.R3.1/HKP.05.00/2017.

## REFERENCES

- Abazari M, Gholamreza Z, Iraj R. 2013. Antimicrobial potentials of *Leptolyngbya* sp. and its synergistic effects with antibiotics. *Jundishapur Journal of Microbiology*. **6**:1—6.
- Bold, H.C., Wynne M.J. 1985. *Introduction to the algae* structure and reproduction. Prentice-Hall, Inc., Englewood Cliffs, New Jersey: xiv+706 pp.
- Beetul K, Sadally SB, Taleb-Hossenkhan N, Bhagooli R, Puchooa D. 2014. An investigation of biodiesel production from microalgae found in Mauritian waters. *Biofuel Research Journal*. **2**:58—64.
- Gerloff-Elias A, Elly S, Thomas P. 2005. Effect of external pH on the growth, photosynthesis and photosynthetic electron transport of *Chlamydomonas acidophila* Negoro, isolated from and extremely acidic lake. *Plant, Cell and Environment.* **28**:1218—1229.
- Hogg S. 2005. Essential Microbiology, 1st ed. John Wiley

- and Sons Ltd, USA: 468 pages.
- Khemka A, Meenu S. 2015. Phycoremediation of dairy wastewater coupled biomass production using *Leptolyngbya* sp. *Journal of Environmental Sciences and Water Resources*. 2: 104—111.
- Komarek J. 2007. Phenotype diversity of the cyanobacterial genus *Leptolyngbya* in the maritime Antarctic. *Polish polar research*. **3**:211—231.
- Lee, R.E. 2008. *Phycology*. Cambridge University Press, New York: ix + 534 pp.
- Madigan M, Martino J, Bender K, Buckley D, Stahl D. 2015. *Brock Biology of Microorganisms*, USA: Pearson education.
- Markou G, Vandamme D, Muylaert K. 2014. Microalgal and cyanobacterial cultivation: the supply of nutrients. *Water research*. **65**:186—202.
- Muster P, Binder A, Schneider K, Bachofen R. 1983. Influence of Temperature and pH on the growth of the thermopilic cyanobacteria *Mastigocladus laminosus* in continous culture. *Plant & Cell Physiology*. **24**:273—280.
- NIES-collection. 2007. *List of strains: Microalgae and protozoa* 7th ed. Nissei Eblo Co., Ltd. Tsukuba: v + 159 pp.
- Olsson-francis K, Simpson AE, Wolff-Boenisch, Cockel CS. 2012. The effect of rock composition on cyanobacterial weathering of crystalline basalt and rhyolite. *Geobiology*. **10**:434—444.
- Prihantini NB. 2015. Polyphasic taxonomy of culturable cyanobacteria isolated from hot springs in west java, Indonesia [dissertation]. Depok (ID): Universitas Indonesia.
- Scholnick S, Nir K. 2006. Metal homeostasis in cyanobacteria and chloroplasts. Balancing benefits and risks to the photosynthetic apparatus. *Plant Physiology*. **141**:805—810.
- Van Den Hoek C, Mann DG, Jahns HM. 2002. Algae: An introduction to phycology, USA: Cambridge university press.